

Immunological Activity of Larch Arabinogalactan and Echinacea: A Preliminary, Randomized, Double-blind, Placebo-controlled Trial

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Abstract

OBJECTIVE: The immunomodulating effects of two Echinacea species, *E. purpurea* and *E. angustifolia* and larch arabinogalactan extracted from *Larix occidentalis* were examined in a randomized, double-blind, placebo-controlled, prospective four-week clinical trial at a naturopathic medical school research center. **SUBJECTS/MATERIALS:** Forty-eight healthy female volunteers (22-51 y) were randomly assigned to one of six groups: standardized extract of *E. purpurea* (EP); ultra-refined *E. purpurea/E. angustifolia* (urEPA); *E. purpurea/E. angustifolia* (EPA); *E. purpurea/E. angustifolia* plus larch arabinogalactan (EPALA); larch arabinogalactan (LA); or placebo. **METHODS:** Immunological tests with enumerative measurements, stool cultures for *Lactobacillus acidophilus* and yeast, and health-related quality of life (HRQoL) using the Medical Outcomes Study derived SF-36 self-administered questionnaire were assessed at baseline and at four weeks. **RESULTS:** Complement properdin increased by 21 percent in the EPA group ($p < 0.05$) and by 18 percent in the EPALA group ($p < 0.05$), compared to the placebo group ($p > 0.05$). SF-36 showed improvements in overall physical health, vitality, and emotional health in the same two groups (EPA and EPALA). **DISCUSSION:** Volunteers in the EPA and EPALA groups had increased production of complement properdin after four weeks of intervention. The increased

complement properdin may be an indication of one aspect of immune system stimulation in patients treated with either *E. purpurea/E. angustifolia* or *E. purpurea/E. angustifolia* plus larch arabinogalactan.

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Introduction

Both *Echinacea purpurea* and *Echinacea angustifolia* appear to activate non-specific cellular and humoral immunity and the complement system.¹⁻⁵ Both of these species stimulate the immune system by increasing the production and activity of leukocytes, lymphocytes, and monocytes, as well as cytokines.⁶⁻¹¹ *E. purpurea* and *E. angustifolia* have been shown to enhance the immune system in both animal models and clinical

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trials.¹²⁻¹⁶ The enhanced immune function and phagocytic activities have been documented with natural killer (NK) cells, macrophages, and neutrophils of the reticuloendothelial system. Cytokine productions of gamma-interferon (IFN- γ), tumor necrosis factor-alpha (TNF- α), interleukin-1 beta (IL-1 β), and interleukin-6 (IL-6) are some of the immune modulators released and stimulated by *E. purpurea* and *E. angustifolia*.¹⁷⁻²⁰ In clinical studies, Echinacea reduced symptoms of common cold, influenza, and acute respiratory infections.²¹⁻²⁴ Randomized controlled trials demonstrate significant reduction in cold symptoms, increased length of time between infections, and more rapid resolution of viral infections.²¹⁻²³ There have been studies that did not find statistically significant improvement in treating the common cold with Echinacea.²⁵ In general, however, extensive studies support the immune-stimulating, antitumor, and anti-inflammatory activities of Echinacea.²⁶⁻³⁰ Echinacea is generally considered to be safe with no significant toxicity or adverse effects.^{31,32}

Constituents of Echinacea include polysaccharides, echinacin, echinacoside, flavonoids, caffeic acid derivatives, essential oils, polyacetylenes, alkylamides, and assorted other chemicals. *E. angustifolia* and *E. purpurea* are the most widely used and extensively studied for their active components in analytical chemistry and clinical application. However, due to the various extraction processes (alcohol, glycerin, standardized extracts, whole plant extracts) and delivery methods (capsule, tablet, tincture, tea) currently available, and the various parts of the Echinacea plant with unknown pharmacodynamics and pharmacokinetics (leaf, stem, flower, root, and their respective biologically active/marker components), it is difficult to determine what form of the botanical is the most effective, safe, and valuable in immune enhancement.³³⁻³⁹

Larch arabinogalactan from *Larix occidentalis* was shown to increase circulating peripheral blood monocytes.⁴⁰ Tumor cells pretreated with larch arabinogalactan enhanced NK cell cytotoxicity and phagocytic capacities of macrophages and lymphocytes, and increased release of various cytokines, such as IFN- γ , TNF- α ,

IL-1 β , and IL-6.⁴⁰⁻⁴⁵ Larch arabinogalactans are a class of long, densely branched high-molecular weight polysaccharides (10,000-120,000 daltons).⁴⁶ High-grade arabinogalactan extracted from *Larix occidentalis* is composed of 90-98 percent arabinogalactan, and experimental analysis has determined larch arabinogalactan to be a highly branched molecule of 3,6-beta-D-galactan.^{46,47} There are numerous patents identified in product development using larch arabinogalactan. According to the Generally Recognized as Safe (GRAS) Notice No. GRN 000047 (FDA, Center for Food Safety & Applied Nutrition, Office of Premarket Approval), functional properties of larch arabinogalactan permit its use as a film-former, foam adhesive, additive, thickener, bulking agent, emulsifier, and as a therapeutic agent. Based on food grade status and numerous studies supporting the safety of larch arabinogalactan, it is considered to be extremely safe with minimum to no toxicity.

Immune-stimulating effects of the combination of Echinacea and larch arabinogalactan have been reported.⁴⁸ Healthy donor blood treated with a combination of larch arabinogalactan, *E. purpurea*, and *E. angustifolia* in 24-hour incubation showed significant increase in macrophage cell density, and the greatest immune cell stimulation and proliferation when compared to single agent vitamins and minerals. The same study showed the combination Echinacea and larch arabinogalactan had a greater immune-enhancing effect than the individual effects of either Echinacea or larch arabinogalactan alone.

Objectives

In this study, the immune-stimulating effects of combination Echinacea and larch arabinogalactan observed in previous *in vitro* studies using healthy donor blood samples were tested in healthy volunteers. A randomized, double-blind, placebo-controlled trial was conducted to address the following: (1) to compare the combination therapy to monotherapies of Echinacea and larch arabinogalactan; (2) to assess immunological outcomes following a treatment period of four weeks; and (3) to assess quality of life outcomes of the

intervention. In this report, the immunomodulating effects of different *E. purpurea* whole herb and *E. angustifolia* root preparations, larch arabinogalactan 90-percent concentration extracted from *Larix occidentalis*, and combination Echinacea and larch arabinogalactan are presented.

Subjects

Subject Population

The clinical trial was approved by the Institutional Review Board (Human Subject Protection Review Committee) of Southwest College of Naturopathic Medicine & Health Sciences. Subjects were selected according to the study inclusion and exclusion criteria (Table 1). Subjects were

recruited with newspaper advertisements and posted announcements. A potential 128 female participants were interviewed in a preliminary screening; of these, 48 were selected to participate and randomly assigned to one of six groups. Only females were included in the study to reduce variability of both outcomes and analysis of results. Subjects with major illness (cancer, diabetes, or cardiovascular or autoimmune/immune diseases), acute illness at enrollment and during study period (upper respiratory tract infections, sinusitis, and other acute infections), or subjects taking any known immune-enhancing or altering supplements or medication were excluded from the study. All subjects were followed at Southwest Naturopathic Medical Center in Scottsdale, Arizona. Each subject provided written, informed consent to participate, and were informed of possible rare transient reactions from taking herbal supplements, such as nausea and stomach discomfort.

Table 1. Clinical and Demographic Data of Subjects

CHARACTERISTIC	No. (%) OF SUBJECTS
Sex	
Female	48 (100)
Race or ethnic group	
Black	2 (4)
Hispanic	1 (2)
White	45 (94)
Non-smoker	48 (100)
Healthy*	
No major illness	48 (100)
No acute illness	48 (100)
Age (yr)	
Mean	36.7
Range	22-51

***Major illness: cancer, diabetes, cardiovascular, autoimmune/immune diseases. Acute illness at enrollment and during study period: upper respiratory tract infections, sinusitis, and other acute infections.**

Evaluation of Subjects

Medical history intake and vital signs were recorded at the beginning of the study. Subjects were interviewed at two weeks to monitor compliance with pill taking and to record any adverse effects or changes in health or daily activities. Lifestyle changes inconsistent with the study requirements during the four-week intervention period were also noted including: excessive alcohol intake, recreational drug or new prescription/non-prescription drug use, strenuous exercise, diet changes, and inclusion of other complementary/alternative therapies that could affect the immune system.

Materials

Intervention Plan

All subjects took daily doses of the supplement on an empty stomach for four weeks, two capsules in the morning and at bedtime. There were six

Table 2. Dose Description

GROUP	INTERVENTION THERAPY	DOSE per day
EP	<i>E. purpurea</i> whole herb extract 4% phenols	1500 mg/d
urEPA	<i>E. purpurea</i> whole herb extract 4% phenols Ultra refined <i>E. purpurea</i> whole herb and <i>E. angustifolia</i> root	780 mg/d 680 mg/d
EPA	<i>E. purpurea</i> whole herb extract 4% phenols <i>E. purpurea</i> whole herb <i>E. angustifolia</i> root	908 mg/d 464 mg/d 36 mg/d
EPALA	<i>E. purpurea</i> whole herb extract 4% phenols <i>E. purpurea</i> whole herb <i>E. angustifolia</i> root Larch arabinogalactan 90%	908 mg/d 464 mg/d 36 mg/d 1500 mg/d
LA	Larch arabinogalactan 90%	1500 mg/d
Placebo	Alfalfa and rice	1500 mg/d

groups in the study (Table 2): (1) *E. purpurea* whole herb extract (4% phenols; 1.5 g/day) (EP); (2) *E. purpurea* whole herb extract (4% phenols; 780 mg/day), ultra-refined *E. purpurea* whole herb, and *E. angustifolia* root (680 mg/day) (urEPA); (3) *E. purpurea* whole herb extract (4% phenols; 908 mg/day), *E. purpurea* whole herb (464 mg/day), and *E. angustifolia* root (36 mg/day) (EPA); (4) *E. purpurea* whole herb extract (4% phenols; 908 mg/day), *E. purpurea* whole herb (464 mg/day), *E. angustifolia* root 36 mg/day, and larch arabinogalactan (90%; 1.5 g/day) (EPALA); (5) larch arabinogalactan (90%; 1.5 g/day) (LA); or (6) placebo (alfalfa and rice flour; 1.5 g/day). Subjects were compliant and followed study instructions during the four weeks, avoiding new alternative therapies, dietary supplements,

and excessive alcohol (Table 3). All capsules were indistinguishable in size, color, and taste. Celestial Seasonings, Inc. and Larex, Inc. provided the investigational supplements and placebo for the study.

Intervention Analytical Chemistry

The *E. purpurea* whole herb extract (4% phenols), *E. purpurea* whole herb, *E. angustifolia* root, and larch arabinogalactan used in the study were extracted from the same lot numbers. The active components of the Echinacea preparations were analyzed for percent phenols and microbial characteristics by Nutritional Laboratories International (Lolo, MT). Microbial analysis showed the herbal preparations were negative for *E. coli* and *Salmonella*.

Table 3. Subject Compliance

STUDY REQUIREMENTS	No. (%) OF SUBJECTS
No new physical therapies during study*	48 (100)
Alcohol < 3 drinks per week during study	48 (100)
Compliance to dosage and pill-taking 100%	
EP	5 (62)
urEPA	8 (100)
EPA	8 (100)
EPALA	8 (100)
LA	8 (100)
Placebo	8 (100)
No new supplements during study†	
EP	8 (100)
urEPA	8 (100)
EPA	8 (100)
EPALA	8 (100)
LA	8 (100)
Placebo	7 (90)

*Physical therapies including acupuncture, chiropractic, and other musculoskeletal/connective tissue therapies.

†Supplements including nutritional, herbal, and other complementary/alternative therapies.

bacterial stool culture for *Lactobacillus acidophilus* and stool fungus culture for yeast in colony forming units per gram (cfu/g). Culture medium selective for gram+ was used for *Lactobacillus acidophilus* and mold-inhibiting medium was used for yeast cultures. The procedure detected colonies with more than 100 organisms per colony (1cfu/g=100 *Lactobacillus acidophilus* or yeast), and colonies of <100 organisms were not reported. ABO blood typing was assessed in all subjects at the beginning of the study. Sonora Quest Laboratory (Phoenix, AZ) and Specialty Laboratory (Santa Monica, CA) performed the enumerative measurements.

Subjective reporting on HRQoL was assessed using the standard SF-36 and Symptoms Specific Assessment (SSA) derived from Medical Outcomes Study (MOS), including gastrointestinal function, sleep pattern, and mood.⁴⁹⁻⁵² These instruments were self-administered by subjects at baseline and at four weeks. The SSA was developed by the investigators to specifically address quality of life effects with respect to gastrointestinal function, sleep pattern, and mood. The SF-36 was chosen for its multi-dimensionality, brevity, and previous successful application in a variety of diseases. Responses to the 36 items on SF-36 assess a number of HRQoL domains, ranging from predominantly social and emotional well-being to overall mental and physical health and vitality.

Methods

Criteria for Response

The effects of the investigational supplements on the subjects' health and immune function were assessed with immunological tests and subjective reporting on quality of life. Vital signs were measured (blood pressure, radial pulse, respiration rate, and temperature), and blood and stool samples were collected at baseline and at four weeks.

Immunological tests with enumerative measurements included: total white blood cell (WBC), neutrophils, lymphocytes, monocytes; NK cell quantitative; complement properdin (CP); TNF- α ; Epstein-Barr Virus viral capsid antigen IgG antibody (EBV VCA IgG Ab); cytomegalovirus IgG antibody (CMV IgG Ab); and aerobic

Table 4. Mean Complement Properdin and TNF- α and their P values

GROUP	COMPLEMENT PROPERDIN			TNF- α		
	BASELINE %	WEEK 4 %	P VALUES	BASELINE pg/mL	WEEK 4 pg/mL	P VALUES
EP	94 + 25	95 + 33	NS	15 + 23	5 + 1	NS
urEPA	71 + 30	94 + 24	NS	8 + 2	5 + 2	0.040
EPA	60 + 30	86 + 24	0.029	14 + 9	7 + 4	NS
EPALA	47 + 27	70 + 25	0.020	15 + 7	8 + 4	0.034
LA	112 + 97	72 + 29	NS	12 + 8	6 + 1	0.044
Placebo	96 + 29	101 + 34	NS	17 + 32	7 + 2	NS

Plus-minus values are means +SD. NS denotes no significance. P values by two-tailed Student's t-test.

Statistical Analysis

The two-tailed Student's *t*-test was used to assess the differences between baseline and end of treatment. The outcomes of the five active

groups and the placebo group were compared between baseline and four weeks. Self-administered SF-36 and SSA were also statistically analyzed for significance in the study groups.

Figure 1. Mean Complement Properdin

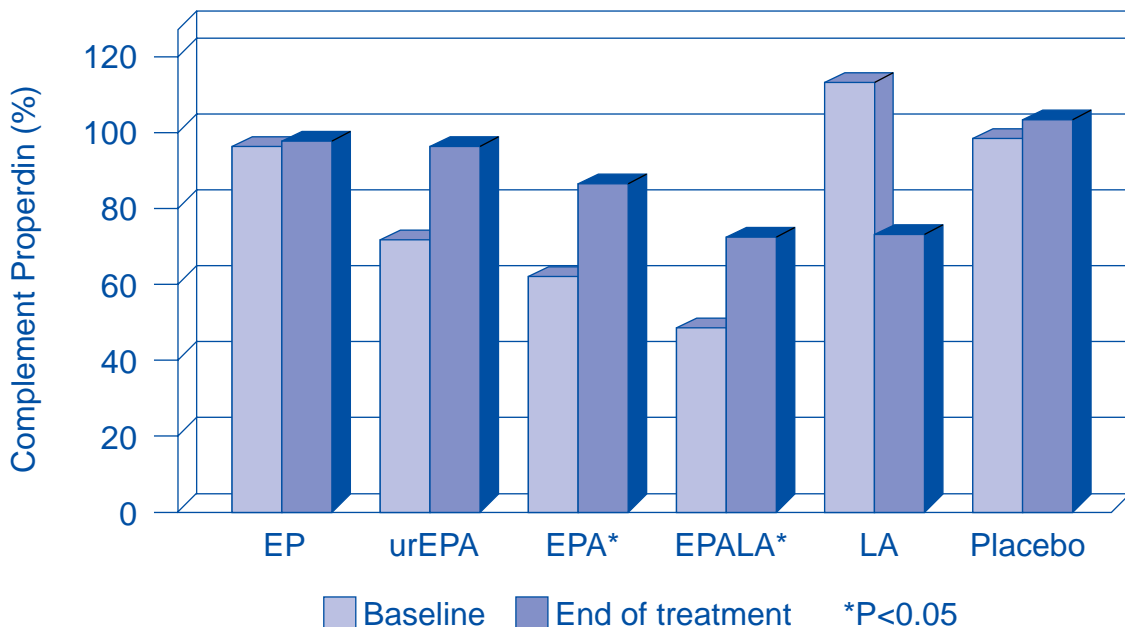


Table 5. Mean Hematological and Immunological Values

GROUP	WBC		NEUTROPHILS		LYMPHOCYTES		MONOCYTES	
	BASELINE m/mm ³	WEEK 4 m/mm ³	BASELINE cells/mm ³	WEEK 4 cells/mm ³	BASELINE cells/mm ³	WEEK 4 cells/mm ³	BASELINE cells/mm ³	WEEK 4 cells/mm ³
EP	5.94 + 1.44	4.86 + 0.90	3569 + 1248	2620 + 851	1881 + 597	1762 + 336	386 + 70	359 + 90
urEPA	5.56 + 1.03	5.06 + 1.08	3336 + 868	2653 + 796	1704 + 284	1785 + 318	402 + 113	380 + 188
EPA	5.88 + 1.77	5.65 + 1.48	3519 + 1510	3364 + 1319	1845 + 438	1811 + 466	407 + 115	354 + 72
EPALA	4.77 + 1.45	4.49 + 0.94	2745 + 1179	2640 + 800	1547 + 308	1408 + 386	354 + 72	318 + 71
LA	6.05 + 1.61	5.71 + 1.36	3829 + 1194	3647 + 1183	1728 + 482	1587 + 586	364 + 55	363 + 75
Placebo	5.75 + 2.10	5.85 + 2.25	3345 + 1306	3469 + 1440	1846 + 725	1852 + 687	375 + 166	362 + 128

Plus-minus values are means +SD.

Results

Lab Parameters

Complement properdin percent concentration increased significantly in the EPA and EPALA groups (Table 4). At four weeks the CP in the EPA group was (mean [±SD]) 86±24 (baseline 60±30) and in EPALA was 70±25 (baseline 47±27), compared with 101±34 (baseline 96±29) in the placebo group (Figure 1). CP difference in the EPA group between baseline and four weeks was 26±27 (21.0% increase) (p=0.029) and in the EPALA group it was 22±19 (18.0% increase) (p=0.020); while in the placebo group it was 5±34 (p=0.687). TNF-α decreased in urEPA, EPALA, and LA groups (p=0.040, p=0.034, and p=0.044, respectively) (Table 4). There were no statistically significant pattern changes in the hematological or other immunological serum chemistry (Table 5). Aerobic bacterial culture for *Lactobacillus acidophilus* and fungus culture for yeast showed no significant results.

Quality of Life Assessment

SF-36 improved in the EPA and EPALA groups at end of treatment (p=0.042 and p=0.031, respectively) (Table 6), and showed enhanced vitality and physical, emotional and mental health. The SSA decreased in the EPA and EPALA groups (p=0.003 and p=0.015, respectively) (Table 6). The decreases in SSA mean scores demonstrate improvement in gastrointestinal function, sleep pattern, and mood (Figure 2).

Subjects also reported increased bowel movements (BM) and changes in stool characteristic (looser quality and larger quantity) in the following frequency: EP, 4 of 8 subjects (50%); urEPA, 3 of 7 (43%); EPA, 4 of 8 (50%); EPALA, 3 of 7 (43%); LA, 6 of 8 (75%); and placebo, 2 of 8 (25%). The changes in BM reports were most dramatic in the larch arabinogalactan group, particularly in the quality of stool consistency. The majority of subjects in this group experienced varying degrees of changes in their BM habits. None of the subjects reported any discomfort and did not discontinue the study due to BM changes.

Adverse Reactions

Two of the 48 subjects experienced adverse reactions two weeks into the study. One volunteer in the urEPA group discontinued due to self-reported anxiety, nervousness, and heart palpitation while taking the supplements. Another volunteer in the EP group reported bilateral arthritic symptoms over her wrist, metacarpophalangeal, and proximal interphalangeal joints; however, the symptoms were similar in location and quality to arthritic symptoms experienced over 10 years previously. The symptoms of the two subjects resolved without complication upon discontinuing the supplement.

Discussion

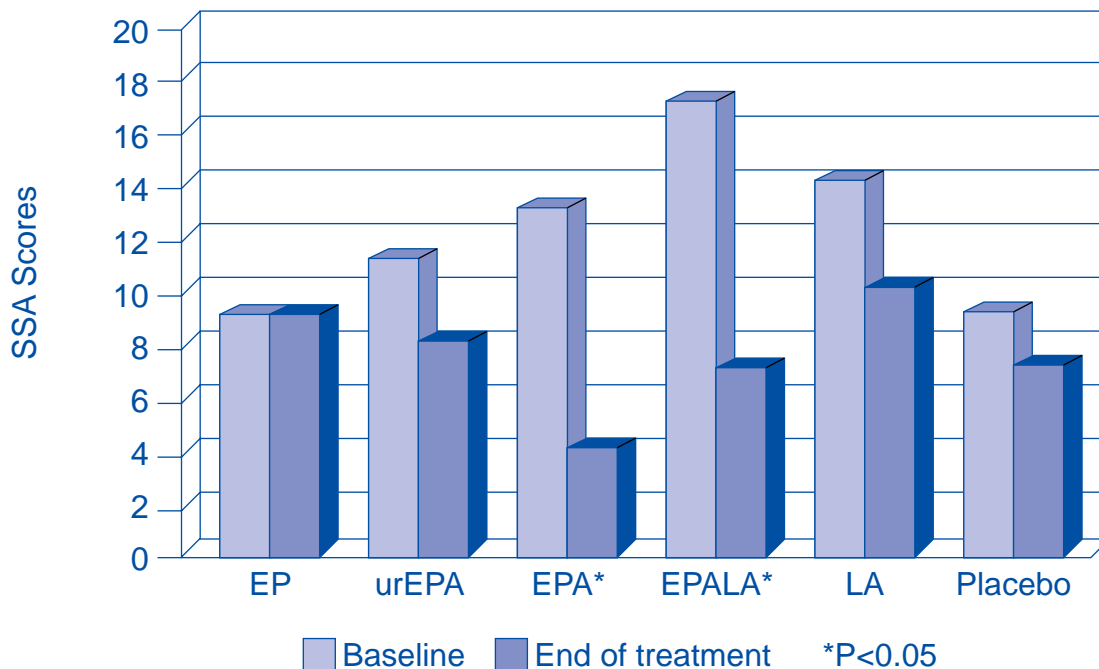
E. purpurea, *E. angustifolia*, and larch arabinogalactan supplements were generally well tolerated. Gastrointestinal function, sleep pattern, mood, and emotional health items of SSA improved

Table 6. P values of SF-36 and Symptoms Specific Assessment

GROUP	SF-36	Symptoms Specific Assessment
EP	NS	NS
urEPA	NS	NS
EPA	0.042	0.003
EPALA	0.031	0.015
LA	NS	NS
Placebo	NS	NS

NS denotes no significance. P values by two-tailed Student's t-test.

Figure 2. Mean Scores of Symptoms Specific Assessment



significantly in the same groups with higher SF-36 scores, which were the EPA and EPALA groups. Based on SF-36 and SSA outcomes related to HRQoL, the most significant benefits to subjects' health and vitality were observed in the EPA and EPALA groups.

The dense polysaccharides of larch arabinogalactan are considered a good source of dietary fiber, improving intestinal microflora such as *Bifidobacterium*, *Lactobacillus acidophilus*, and *Enterobacteriaceae* in human studies.⁵³⁻⁵⁵ Larch arabinogalactan fibers are fermented by gastrointestinal microflora resulting in the production of short-chain fatty acids (especially butyrate and propionate). These fatty acids are of particular value to colonocytes, and are the intestinal epithelial cells' preferred fuel for energy generation. The fiber dense quality of larch arabinogalactan may explain the greater changes in BM from the LA intervention than from the Echinacea formulas or placebo. Although many of the subjects had changes in their digestive habits, no conclusive data pertaining to stool cultures of *Lactobacillus acidophilus* and yeast emerged from the study. Vital signs collected at baseline and four weeks showed no negative or positive correlation with the treatment intervention. No statistical correlations or patterns were observed with blood type and outcomes of the study; the randomization of subjects to the six groups was not matched for blood types at the beginning of the study.

The increase in complement properdin concentrations in the *E. purpurea*/*E. angustifolia* and *E. purpurea*/*E. angustifolia* plus larch arabinogalactan groups may indicate stimulation of the complement immune system. Biological activities associated with complement activation include cell lysis, opsonization, enhanced phagocytosis, complement receptor activation, chemotaxis, activation of neutrophils and monocytes, and clearance of immune complexes.^{56,57} The stimulation by EPA and EPALA may result in enhanced phagocytosis and immune function, and increased production and release of cytokines, which can induce expression of other cytokines and immune cells, further benefiting the specific and non-specific immune system. However, due to the large

variance in the baseline values of the six groups, the outcomes of the study should be considered cautiously in supporting immune effects of the interventions. Thus, TNF- α decreases ($p < 0.05$) in the three groups – urEPA, EPALA, and LA – and complement properdin changes (increased in EPA and EPALA and decreased in LA) (Table 4) do not suggest immunomodulating activities of the intervention in this study and remain suspect, requiring further investigation.

The preliminary nature of the project did not include extensive screening of the subjects prior to enrollment. Testing their WBC and other immune parameters with requisite ranges (upper and lower limits) and examination of their medical records with focused subject selection criteria may have prevented such high variance in the baseline values. The wide age range (22-51); personal/professional background (student, working professional, etc.); diverse stress indicators, diet behaviors, and daily activities of the subjects; and lack of matching based on such diversity may have contributed to the dissimilar lab values. Lack of statistically significant activities of Echinacea and larch arabinogalactan formulas in this study may also be the result of methodology used – subject characteristics, insufficient sample size, and inadequate length of therapy.

This study raises questions leading to future proposals to further explore the immune effects of Echinacea and larch arabinogalactan. Previous clinical trials have shown increased secretion and activity of leukocytes and cytokines (IFN- γ , TNF- α , IL-1 β , and IL-6), while other studies have supported the benefits of Echinacea and larch arabinogalactan in reducing symptoms and recovery time from acute respiratory tract infections, such as the common cold and influenza. Activation of the complement system and increased production of complement properdin may be another immune factor stimulated by Echinacea and combination Echinacea and larch arabinogalactan. Demonstrating clinical efficacy and pharmacodynamics of Echinacea and larch arabinogalactan require more research to understand the components involved in stimulating the natural immune defense system.

In addition to evaluating the levels of complement properdin, measurement of other components of the complement system may provide more conclusive evidence on complement system activation by Echinacea and larch arabinogalactan. The role of Echinacea and larch arabinogalactan to stimulate the specific and non-specific immune system should also be further explored to understand the specific antimicrobial and anti-inflammatory actions of these immunomodulators. Future studies designed to assess immune response to both short- and long-term interventions and antigen-induced immune responses may provide better understanding of these botanical extracts.

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References

- Bauer R, Wagner H. Echinacea species as potential immunostimulatory drugs. *Econ Med Plant Res* 1991;5:253-321.
- Willard T. *Textbook of Advanced Herbology*. Alberta, Canada: Wild Rose College of Natural Healing Ltd; 1992:85-86.
- Blumenthal M, Goldberg A. *The Complete German Commission E Monographs*. Boston, MA: Integrative Medicine Communications; 1998:122-123.
- Murray MT. *The Healing Power of Herbs*, 2nd ed. Rocklin, CA: Prima Publishing; 1995:92-107.
- Murray MT, Pizzorno J. *Encyclopedia of Natural Medicine*, 2nd ed. Rocklin, CA: Prima Publishing; 1998:159-160.
- Sun LZ, Currier NL, Miller SC. The American coneflower: a prophylactic role involving nonspecific immunity. *J Altern Complement Med* 1999;5:437-446.
- Melchart D, Linde K, Worku F, et al. Results of five randomized studies on the immunomodulatory activity of preparations of Echinacea. *J Altern Complement Med* 1995;1:145-160.
- Wagner H, Jurcic K. Immunologic studies of plant combination preparations. *In-vitro* and *in-vivo* studies on the stimulation of phagocytosis. *Arzneimittelforschung* 1991;41:1072-1076. [Article in German]
- Bauer R, Jurcic K, Puhlmann J, Wagner H. Immunologic *in vivo* and *in vitro* studies on Echinacea extracts. *Arzneimittelforschung* 1988;38:276-281. [Article in German]
- Vomel T. Effect of a plant immunostimulant on phagocytosis of erythrocytes by the reticulohistiocytary system of isolated perfused rat liver. *Arzneimittelforschung* 1985;35:1437-1439. [Article in German]
- Rehman J, Dillow JM, Carter SM, et al. Increased production of antigen-specific immunoglobulins G and M following *in vivo* treatment with the medicinal plants *Echinacea angustifolia* and *Hydrastis canadensis*. *Immunol Lett* 1999;68:391-395.
- Steinmuller C, Roesler J, Grottrup E, et al. Polysaccharides isolated from plant cell cultures of *Echinacea purpurea* enhance the resistance of immunosuppressed mice against systemic infections with *Candida albicans* and *Listeria monocytogenes*. *Int J Immunopharmacol* 1993;15:605-614.
- Bukovsky M, Vaverkova S, Kostalova D, Magnusova R. Immunomodulating activity of ethanol-water extracts of the roots of *Echinacea gloriosa* L., *Echinacea angustifolia* DC, and *Rudbeckia speciosa* Wenderoth tested on the immune system in C57BL6 inbred mice. *Cesk Farm* 1993;42:184-187. [Article in Slovak]
- Roesler J, Steinmuller C, Kiderlen A, et al. Application of purified polysaccharides from cell cultures of the plant *Echinacea purpurea* to mice mediates protection against systemic infections with *Listeria monocytogenes* and *Candida albicans*. *Int J Immunopharmacol* 1991;13:27-37.
- Bukovsky M, Kostalova D, Magnusova R, Vaverkova S. Testing for immunomodulating effects of ethanol-water extracts of the above-ground parts of the plants Echinacea (Moench) and Rudbeckia L. *Cesk Farm* 1993;42:228-231. [Article in Slovak]
- Wildfeuer A, Mayerhofer D. The effects of plant preparations on cellular functions in body defense. *Arzneimittelforschung* 1994;44:361-366. [Article in German]

17. Roesler J, Emmendorffer A, Steinmuller C, et al. Application of purified polysaccharides from cell cultures of the plant *Echinacea purpurea* to test subjects mediates activation of the phagocyte system. *Int J Immunopharmacol* 1991;13:931-941.
18. Luettig B, Steinmuller C, Gifford GE, et al. Macrophage activation by the polysaccharide arabinogalactan isolated from plant cell cultures of *Echinacea purpurea*. *J Natl Cancer Inst* 1989;81:669-675.
19. Coeugniet EG, Elek E. Immunomodulation with *Viscum album* and *Echinacea purpurea* extracts. *Onkologie* 1987;10:27-33.
20. Stimpel M, Proksch A, Wagner H, Lohmann-Matthes ML. Macrophage activation and induction of macrophage cytotoxicity by purified polysaccharide fractions from the plant *Echinacea purpurea*. *Infect Immun* 1984;46:845-849.
21. Brinkeborn RM, Shah DV, Degenring FH. Echinaforce and other *Echinacea* fresh plant preparations in the treatment of the common cold. A randomized, placebo-controlled, double-blind clinical trial. *Phytomedicine* 1999;6:1-6.
22. Braunig B, Dorn M, Knick E. *Echinacea purpurea* radix for strengthening the immune response in flu-like infections. *Z Phytother* 1992;13:7-13.
23. Schoneberger D. The influence of immune-stimulating effects of pressed juice from *Echinacea purpurea* on the course and severity of colds. Results of double-blind study. *Forum Immunol* 1992;8:2-12.
24. Dorsch W. Clinical application of extracts of *Echinacea purpurea* or *Echinacea pallida*. Critical evaluation of controlled clinical studies. *Z Arztl Fortbild* 1996;90:117-122. [Article in German]
25. Grimm W, Muller HH. A randomized controlled trial of the effect of fluid extract of *Echinacea purpurea* on the incidence and severity of colds and respiratory infections. *Am J Med* 1999;106:138-143.
26. Lersch C, Zeuner M, Bauer A, et al. Non-specific immunostimulation with low doses of cyclophosphamide (LDCY), thymostimulin, and *Echinacea purpurea* extracts (echinacin) in patients with far advanced colorectal cancers: preliminary results. *Cancer Invest* 1992;10:343-348.
27. Muller-Jakic B, Breu W, Probstle A, et al. *In vitro* inhibition of cyclooxygenase and 5-lipoxygenase by alkamides from *Echinacea* and *Achillea* species. *Planta Med* 1994;60:37-40.
28. Tragni E, Galli CL, Tubaro A, et al. Anti-inflammatory activity of *Echinacea angustifolia* fractions separated on the basis of molecular weight. *Pharmacol Res Commun* 1988;20:87-90.
29. Tragni E, Tubaro A, Melis S, Galli CL. Evidence from two classic irritation tests for an anti-inflammatory action of a natural extract, Echinacina B. *Food Chem Toxicol* 1985;23:317-319.
30. Tubaro A, Tragni E, Del Negro P, et al. Anti-inflammatory activity of a polysaccharidic fraction of *Echinacea angustifolia*. *J Pharm Pharmacol* 1987;39:567-569.
31. Miller LG. Herbal medicinals: selected clinical considerations focusing on known or potential drug-herb interactions. *Arch Intern Med* 1998;158:2200-2211.
32. Mengs U, Clare CB, Poiley JA. Toxicity of *Echinacea purpurea*. Acute, subacute and genotoxicity studies. *Arzneimittelforschung* 1991;41:1076-1081.
33. Gaisbauer M, Schleich T, Stickl HA, Wilczek I. The effect of *Echinacea purpurea* Moench on phagocytosis in granulocytes measured by chemiluminescence. *Arzneimittelforschung* 1990;40:594-598.
34. Schumacher A, Friedberg KD. The effects of *Echinacea angustifolia* on non-specific cellular immunity in the mouse. *Arzneimittelforschung* 1991;41:141-147. [Article in German]
35. Bauer R. *Echinacea* drugs-effects and active ingredients. *Z Arztl Fortbild* 1996;90:111-115. [Article in German]
36. Wagner H, Proksch A, Riess-Maurer I, et al. Immunostimulating action of polysaccharides (heteroglycans) from higher plants. *Arzneimittelforschung* 1985;35:1069-1075. [Article in German]
37. Bone K. *Echinacea*: When should it be used? *Altern Med Rev* 1997;2:451-458.
38. Schulthess BH, Giger E, Baumann TW. *Echinacea*: anatomy, phytochemical pattern, and germination of the achene. *Planta Med* 1991;57:384-388.

39. Egert D, Beuscher N. Studies on antigen specificity of immunoreactive arabinogalactan proteins extracted from *Baptisia tinctoria* and *Echinacea purpurea*. *Planta Med* 1992;58:163-165.
40. Hauer J, Anderer FA. Mechanism of stimulation of human natural killer cytotoxicity by arabinogalactan from *Larix occidentalis*. *Cancer Immunol Immunother* 1993;36:237-244.
41. Kiyohara H, Cyong JC, Yamada H. Relationship between structure and activity of an anti-complementary arabinogalactan from the roots of *Angelica acutiloba* Kitagawa. *Carbohydr Res* 1989;193:193-200.
42. Shimizu N, Tomoda M, Gonda R, et al. The major peptic arabinogalactan having activity on the reticuloendothelial system from the roots of rhizomes of *Saposhnikovia divaricata*. *Chem Pharm Bull* 1989;37:1329-1332.
43. Hayashida Y, Kurimoto S, Yamamoto N. Effects of lymphokine-activated killer cells on human retinoblastoma cells (Y-79) *in vitro*: enhancement of the activity by a polysaccharide preparation, Krestin. *Biochem Biophys Res Commun* 1991;174:107-114.
44. Gonda R, Tomoda M, Ohara N, Takada K. Arabinogalactan core structure and immunological activities of ukonon C, an acidic polysaccharide from the rhizome of *Curcuma longa*. *Biol Pharm Bull* 1993;16:235-238.
45. Causey JL, Robinson RR, Feirtag JM, et al. Effects of larch arabinogalactan on human peripheral blood mononuclear cells: results from *in vivo* and *in vitro* human trials. Dept of Food Science and Nutrition, University of Minnesota; St. Paul, MN: 1999. Unpublished.
46. Kelly GS. Larch arabinogalactan: clinical relevance of a novel immune-enhancing polysaccharide. *Altern Med Rev* 1999;4:96-103.
47. D'Adamo P. Larch arabinogalactan. Research report. *J Naturopathic Med* 1990;6:33-37.
48. Causey J. *In vitro* macrophage cell proliferation study. Dept of Food Science and Nutrition, University of Minnesota; St. Paul, MN: 1999. Unpublished.
49. Ware JE Jr, Sherbourne CD. The MOS 36-item short-form health survey (SF-36). I. Conceptual framework and item selection. *Med Care* 1992;30:473-483.
50. McHorney CA, Ware JE Jr, Lu JF, Sherbourne CD. The MOS 36-item Short-Form Health Survey (SF-36): III. Tests of data quality, scaling assumptions and reliability across diverse patient groups. *Med Care* 1994;32:40-66.
51. McHorney CA, Ware JE Jr, Raczek AE. The MOS 36-Item Short-Form Health Survey (SF-36): II. Psychometric and clinical tests of validity in measuring physical and mental health constructs. *Med Care* 1993;31:247-263.
52. Ware J, Snow K, Kosinski M, Gandek B. *SF-36 Health Survey Manual and Interpretation Guide*. Boston, MA: New England Medical Center, The Health Institute; 1993.
53. Crociani F, Alessandrini A, Mucci MM, Biavati B. Degradation of complex carbohydrates by *Bifidobacterium* spp. *Int J Food Microbiol* 1994;24:199-210.
54. Robinson RR, Feirtag J, Slavin JL. Effects of dietary arabinogalactan on gastrointestinal and blood parameters in healthy human subjects. *J Am Coll Nutr* 2001;20:279-285.
55. Salyers AA, Arthur R, Kuritza A. Digestion of larch arabinogalactan by a strain of human colonic *Bacteroides* growing in continuous culture. *J Agric Food Chem* 1981;29:475-480.
56. Berkow R. *Merck Manual*, 16th ed. Rahway, NJ: Merck Research Laboratories; 1992:296-302.
57. Guyton AC. *Textbook of Medical Physiology*, 8th ed. Philadelphia, PA: W.B. Saunders Co; 1990:374-382.